

Short Paper

p-cresol and oleic acid as reliable biomarkers of estrus: evidence from synchronized Murrah buffaloes

Muniasamy, S.^{1,2}; Muthu Selvam, R.¹; Rajanarayanan, S.^{1,3};
Ramesh Saravanakumar, V.⁴ and Archunan, G.^{1*}

¹Centre for Pheromone Technology, Department of Animal Science, Bharathidasan University, Tiruchirappalli-620 024, India; ²Department of Microbiology, Ayya Nadar Janaki Ammal College, Srivilliputhur, Sivakasi-626 123, India; ³Department of Biotechnology, St. Michael College of Engineering and Technology, Kalayarkovil-630 551, India; ⁴Department of Livestock Production and Management, Veterinary College and Research Institute, Namakkal-600 051, India

*Correspondence: G. Archunan, Centre for Pheromone Technology, Department of Animal Science, Bharathidasan University, Tiruchirappalli-620 024, India. E-mail: garchu56@yahoo.co.in

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Summary

Successful reproductive management in buffaloes depends on effective estrus detection. Urinary pheromones identified from natural estrous cycle have been reported to decipher estrus phase. However, its presence has not been analyzed in the urine after synchronization. Thus, our present investigation was to investigate the influence of synchronized estrus urine in bulls and to examine the presence of estrus-specific compounds ascertained in natural estrus in synchronized buffaloes. Mid-stream urine was collected from six synchronized buffaloes during various phases of estrous cycle and volatiles were examined using GC-MS. Sexual provocation in bulls was established by displaying persistent flehmen and frequent mounting towards estrus urine from synchronized animals. Totally forty-two volatile compounds were identified from three phases of estrous cycle, more specifically 4-methyl phenol (p-cresol) and 9-octadecenoic acid (oleic acid) in estrus urine of synchronized animal as similar to natural estrus. Hence, these chemical cues in buffalo urine might be employed as potential marker candidates for the development of an estrus detection aid.

Key words: CIDR, Flehmen, Mating, 4-methyl phenol, 9-octadecenoic acid

Introduction

Low reproduction potential of buffaloes has been a major concern for decades, which streamed towards a substantial economic loss to developing countries. Protocols have been developed to precisely control the time of ovulation (synchronization) by exogenous hormone administration (De Rensis and López-Gatius, 2007; Perera, 2011). However, poorly timed artificial insemination (AI) due to failure in estrus detection would lead to low rate of conception. Pheromones present in the body fluids of female during estrus elicit behavioral changes in males (Halpin, 1986; Vandenbergh, 1999; Rekwot, 2001; Brennan and Keverne, 2004; Archunan, 2009). Volatiles such as 4-methyl phenol and 9-octadecenoic acid were identified in urine during natural estrous cycle as estrus-specific (Archunan *et al.*, 2014). However, no report is available on urinary volatiles during induced estrous cycle. Hence, it is necessary to confirm the presence of estrus-specific compounds in synchronized buffaloes to prove it as a promising candidate for estrus detection.

Therefore, the present investigation was carried out (i) to study the bull's reproductive behavior towards the urine of synchronized animals and (ii) to examine the urinary volatile compounds in synchronized buffaloes to confirm the presence of estrus-specific compounds

ascertained during natural estrus.

Materials and Methods

Animal and synchronization

Six healthy heifer Murrah buffalo (*Bubalus bubalis*), approximately 20-30 months old, at VCRI, Nammakkal, Tamilnadu, India were recruited for analysis. Animals were housed in sheds and paddocks, and fed with standard diet and water *ad libitum*. All animals were subjected to synchronization of estrus by controlled internal drug releasing (CIDR) kit (EAZI BREED CIDR 1380 kit, Pfizer limited, India). The vaginal insert, containing 1.38 g of progesterone, was inserted (day 1) into the cervico-vaginal region and 5 ml of PGF2 α was injected intramuscularly followed by the removal of kit at the end of the 8th day. On the next day (day 9), 0.75 mg estradiol benzoate was administered (Tauck *et al.*, 2007).

Sample collection

The phases of estrous cycle were comprehended as pre-estrus (days 1-9), estrus (days 11-12), and post-estrus (days 15-18) based on CIDR treatment protocol. The mid-stream urine samples were collected from synchronized buffaloes by free-catch method in sterilized glass bottles and categorized according to estrous cycle

phases (Selvam *et al.*, 2016). One part of each urine sample was used for behavior assay and another part was filtered through nylon mesh (60-120 μm) immediately after collection and stored as aliquots at -20°C .

Behavioral assay

Urine sample collected during different phase of the estrous cycle from all the six animals was sprayed individually on the genital region of dummy buffaloes and bulls were allowed to sniff it for 15 min. The duration of flehmen behavior and the frequency of mounting exhibited by the bulls in response to the urine samples were recorded (Rajanarayanan and Archunan, 2004). The genital region was washed cleanly after every assay and a lag-time of 30 min was given between two assays. All experiments were performed in triplicate. Statistical analysis was performed using Microsoft excel and results were recorded in mean \pm SEM.

Sample preparation for GC-MS

Test urine sample was mixed with DCM (1:1) and filtered through a silica-gel column (60-120 mesh) for 30 min at RT. The filtered extract was reduced to 1/5 of its original volume by cooling with liquid nitrogen. The sample was fractionated and subjected to GC-MS analysis (QP-5050, Shimadzu, Japan) as per Rajanarayanan and Archunan (2011). The identified estrus-specific compounds were then compared with the standard compounds run under the same conditions, and confirmed on the basis of retention time shown in GC-MS.

Results

Estrus urine from the synchronized animals induced extended flehmen behavior (24.00 ± 1.31 s), whereas the duration of flehmen decreased when exposed to pre-estrus (14.50 ± 0.53 s) and post-estrus (7.64 ± 2.19 s) urine (Fig. 1A). Similarly, highest frequency of mounting behavior was observed with synchronized estrus urine (7.97 ± 0.40), when compared to pre-estrus (2.61 ± 0.61) and post-estrus (1.97 ± 0.44) urine samples (Fig. 1B).

The chromatogram of synchronized buffalo urine exhibited totally 42 different volatile compounds (Figs. 2A-C) and the existence of each volatile, along with its nature, was reported in Table 1 for each phase of synchronized estrous cycle. Certain volatiles such as 1-chloro-1-buten-3-yne, 1,4-dichloromethane, 2-octanone, 2-ethyl-5-methyl-1,2,3-dioxoborolan-4-one, ethyl-2-methylcyclopentane, methylheptane, diisooctylester-1,2, benzene dicarboxylic acid, and 3-methyl-1-pentanol were present in all phases.

Nevertheless, the compounds viz. methylpyrrolidine, 3-aminoheptane, 2,3-dimethylheptane, 3-methyl-1-pentene, hexadecanol, 2-octyldodecan-1-ol, 5-methyl-4-decene, and 2-methyleicosane were present in pre-estrus and estrus urine; 2-Methyl-2-propenamide, 4-pentanol, 7-methyl-4-undecane, 4-methyl-3-heptanone, 2-methyl-n-phenyl-2-propenamide, N-N-bis(2-hydroxyethyl)-

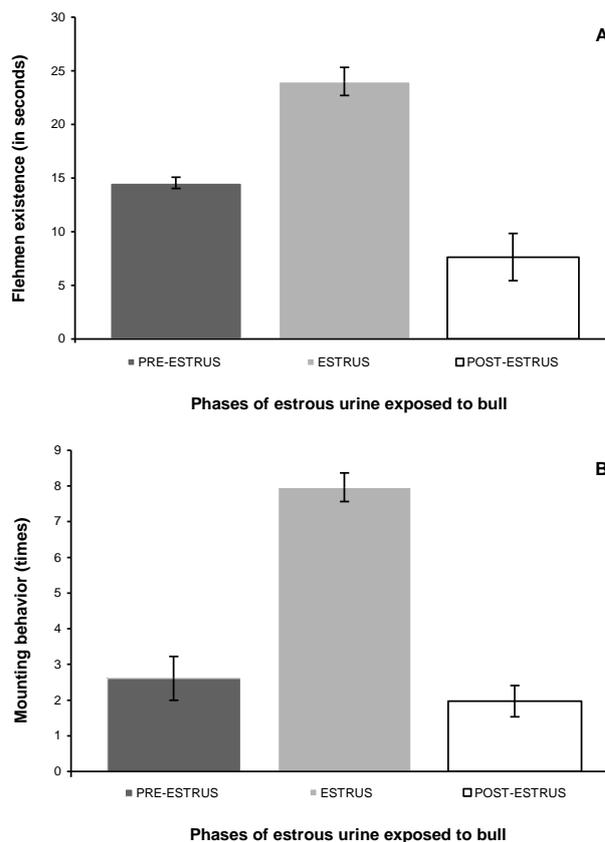
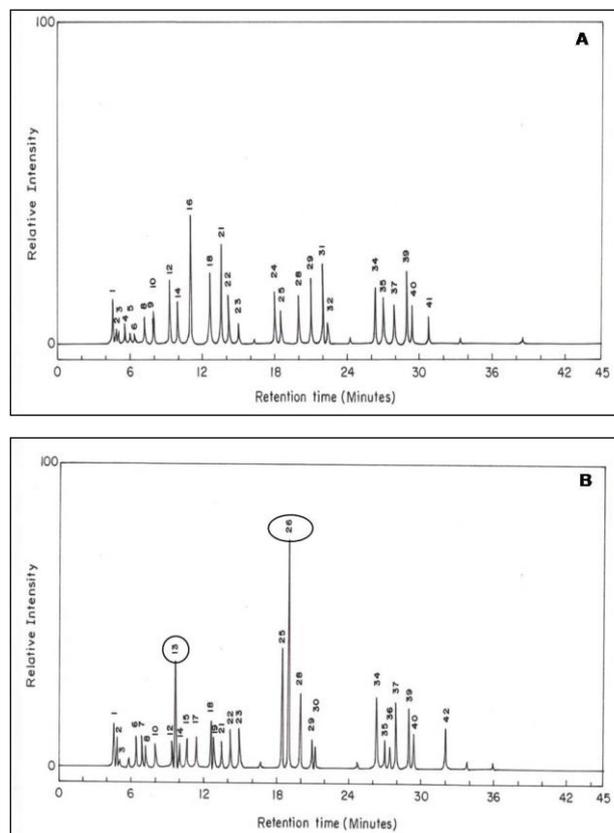


Fig. 1: Behavioral changes of bull towards female buffalo's urine during various phases of estrous cycle. Pane A shows the time spent in exhibiting flehmen behavior and panel B shows the frequency of mounting



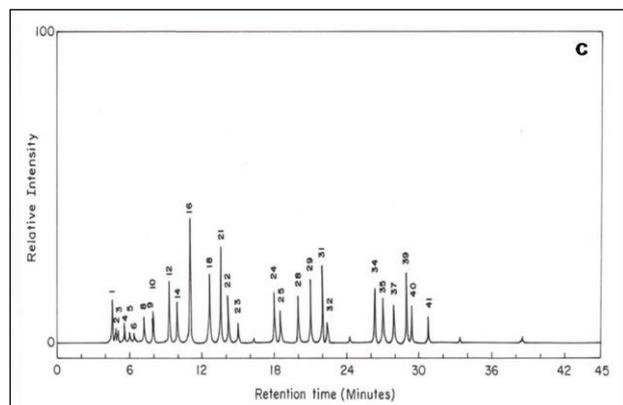


Fig. 2: Representative chromatogram of urine from synchronized animal during various phases of estrous cycle. Panel A: pre-estrus, B: estrus, and C: post-estrus. Peak No. 13 and 26 correspond to p-cresol and oleic acid, respectively

dodecanamide, nonane, decanoic acid, dimethylundecane, tetradecanoic acid, 9-methyl-(z)-5-undecene, and 1-tetradecanol were present in estrus and post-estrus urine; 5-methyl-2-(methylethyl)-

cyclohexanol was identified in pre-estrus and post-estrus urine.

The volatiles such as phenol, 3-propylphenol, 9-octadecenal, and 3-methylbutanol were found only in pre-estrus urine and 1-nitro-2-methylpropane, 2,3-dihydro-4-methylfuran, 3-ethyl-2-methylhexane, 1-chlorotetradecane, hexadecanoic acid, isocyanobutane, and undecanol were identified only in post-estrus urine. However, 4-methyl phenol and 9-octadecenoic acid were the volatile found only during estrus.

Discussion

The phase of estrous cycle was reconfirmed by adopting gynaeco-clinical and biochemical parameters, and coincided with CIDR based characterization. It is well known that estrus urine generally has behaviorally important chemical signals that provoke bulls to manifest pre-mating behaviors (Rivard and Klemm, 1990; Rasmussen, 1998; Rekwot *et al.*, 2001). Estrus signaling compound from urine was suggested to lead bulls to perform flehmen reaction, and later to a more intense

Table 1: List of compounds identified in the urine of female buffaloes by GC-MS

Peak No.	Compound name	PE	E	POE	Mol. Wt	Group
1	1-Chloro-1-buten-3-yne	√	√	√	86	Alkyne
2	2-Methyl-2-propenamide	×	√	√	85	Amide
3	1,4-Dichloromethane	√	√	√	146	Aromatic
4	1-Nitro-2-methylpropane	×	×	√	101	Alkane
5	2,3-Dihydro-4-methylfuran	×	×	√	84	Heterocyclic
6	2-Ethyl-5-methyl-1,2,3-dioxoborolan-4-one	√	√	√	128	Aldehyde
7	Methylpyrrolidine	√	√	×	84	Alkaloid
8	4-Pentanol	×	√	√	86	Alcohol
9	3-Ethyl-2-methylhexane	×	×	√	128	Alkane
10	7-Methyl-4-undecane	×	√	√	168	Alkane
11	Phenol	√	×	×	94	Phenol
12	4-Methyl-3-heptanone	×	√	√	128	Alkane
13	4-Methyl phenol (p-cresol)	×	√	×	108	Phenol
14	2-Octanone	√	√	√	128	Ketone
15	3-Aminoheptane	√	√	×	115	Alkane
16	5-Methyl-2-(methylethyl)-cyclohexanol	√	×	√	156	Alcohol
17	2,3Dimethylheptane	√	√	×	128	Alkane
18	3-Methyl-1-pentanol	√	√	√	102	Alcohol
19	3-Methyl-1-pentene	√	√	×	84	Alkene
20	3-Propylphenol	√	×	×	136	Phenol
21	2-Methyl-n-phenyl-2-propenamide	×	√	√	161	Amide
22	Dimethylundecane	×	√	√	184	Alkane
23	Decanoic acid	×	√	√	172	Acid
24	1-Chlorotetradecane	×	×	√	232	Alkane
25	N-N-Bis(2-hydroxyethyl)-dodecanamide	×	√	√	287	Amide
26	9-octadecenoic acid (oleic acid)	×	√	×	282	Acid
27	Hexadecanol	√	√	×	242	Alcohol
28	Tetradecanoic acid	×	√	√	228	Acid
29	Nonane	×	√	√	128	Alkane
30	2-Octyldodecan-1-ol	×	√	×	298	Alcohol
31	Hexadecanoic acid	×	×	√	256	Acid
32	Isocyanobutane	×	×	√	83	Alkane
33	9-Octadecenal	√	×	×	266	Aldehyde
34	Diisooctylester-1,2-benzenedicarboxylic acid	√	√	√	530	Acid
35	Ethyl-2-methylcyclopentane	√	√	√	112	Alkane
36	5-Methyl-4-decene	√	√	×	154	Alkene
37	Methylheptane	√	√	√	114	Alkane
38	3-Methylbutanol	√	×	×	86	Alcohol
39	9-Methyl-(z)-5-undecene	×	√	√	168	Alkene
40	1-Tetradecanol	×	√	√	214	Alcohol
41	Undecanol	×	×	√	172	Alcohol
42	2-Methyleicosane	√	√	×	296	Alkane

mounting behavior (Garcia *et al.*, 1986). Flehmen was observed with all the tested samples, but the duration (seconds) varied between estrous cycle phases (Fig. 1A). In concordance with previous study (Rajananarayanan and Archunan, 2004), intermittent flehmen was observed in response to estrus urine in the present study, which clearly shows the perceptivity of the bull towards the chemical cues released from female buffaloes. Bulls translate the olfactory cues to the brain to recognize an apt-phase for paring-off (Masaki and Otha, 1990). The high frequency of mounting in response to estrus urine revealed that the volatile chemicals in estrus urine have an important role in mating.

Along with the three reported volatiles in the pre-estrus urine of natural estrous cycle, 3-methylbutanol is additionally present in the pre-estrus urine of synchronized estrous cycle. The estrus-specific volatiles compounds identified in the natural estrous cycle such as p-cresol and oleic acid were also found in the estrus urine of synchronized buffaloes. Cresols have germicidal property and may act as antiseptics, protecting the genitals of both sexes during the sexually active periods. In mares, increased signal of p-cresols during estrus suggested it as an estrus indicating pheromone (Morris *et al.*, 1979; Mozuraitis *et al.*, 2011).

The present study demonstrates that synchronized estrus female influences reproductive behavior in the bull and evidenced that pheromones are indeed present in the synchronized estrus urine as it exists in natural estrus. In addition, the present study convincingly shows that the females could be hormonally regulated to exhibit estrus through synchronization process and the characteristic estrus-specific volatile compounds released during normal estrous cycle are also expressed in synchronized buffaloes. Thus, the bulls displayed significant flehmen, and mounting behavior towards the compounds present during estrus urine in synchronized buffaloes. Based on behavioral observations, the present study concludes p-cresol and oleic acid are the two volatile compounds of estrus phase involved in influencing the sexual arousal and reproductive behavior of bull. Therefore, the study provides strong support to proceed further in developing an estrus detection kit with the estrus-specific volatiles.

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Conflict of interest

No potential financial or personal conflicts of interest

exist between authors.

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