Short Paper

Comparative efficacy of serological diagnostic methods and evaluation of polymerase chain reaction for diagnosis of bovine brucellosis

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Summary

The aim of present study was to compare the diagnostic performance of the different Brucella abortus antigen based serological and molecular tests such as Rose Bengal Plate Test (RBPT), indirect enzyme linked immunosorbent assay (I-ELISA) and polymerase chain reaction (PCR). Out of 89 samples, 44 were positive by I-ELISA, 18 by RBPT and 21 by PCR. Substantial agreement was observed between PCR and I-ELISA (κ=0.48). A slight degree of agreement was observed between RBPT and I-ELISA and PCR (κ=0.18) and RBPT and PCR (κ=0.11). Indirect enzyme linked immunosorbent assay detected more samples as positive among these tests. In conclusion, I-ELISA can be routinely used for an accurate and efficient diagnosis of Brucella infection, because the chances of non-detection of an infected animal in I-ELISA are minimal. However, PCR could be used as a supplement and complement test along with I-ELISA for identification and differentiation of bovine brucellosis.

Key words: Bovine Brucellosis, I-ELISA, PCR, RBPT

Introduction

Brucellosis, caused by Brucella species, is a serious zoonosis affecting a wide range of domestic and wild animals all over the world (Kumar et al., 2009, 2016; Neha et al., 2014; Verma et al., 2014). At present, brucellosis is usually diagnosed based on serological and microbiological tests. Serological methods such as Rose Bengal Plate Test (RBPT) and enzyme linked immunosorbent assay (ELISA) are not always sensitive or specific due to cross-reactivity with other bacterial antigens (OIE, 2012). Isolation of etiological agent is considered as gold standard test in diagnosis of disease, but isolation of Brucella spp. is tedious, time consuming and difficult due to the intra-cellular and fastidious nature of the bacteria and is relatively difficult under field conditions due to its zoonotic nature (Kaynak-Onurdag et al., 2016). Recently, new molecular techniques like the polymerase chain reaction (PCR) have been developed for the detection of Brucella DNA in body fluids having low number of non-viable Brucella organisms (Çiftci et al., 2017). Therefore, the present study was conducted to evaluate PCR assay for the detection of Brucella DNA in serum and from bovine and to compare its performance with serological methods.

Materials and Methods

Sample collection

The present cross-sectional study was carried out in different districts viz., Agra, Amroha, Baghapat, Bulandsahar, Etawah, Firozabad, Ghaziabad, Hapur, Mainpuri and Mathura of the western part of Uttar Pradesh, India during 2014-2015 using a convenient sampling procedure with records of animal determinants. All the animals were handled as per the guidelines of Ethical Committee. 3-5 ml blood samples were aseptically collected from the selected 89 animals by jugular vein-puncture using vaccutainers (BD, USA) and serum was separated by centrifugation and stored at -20°C till tested. A total of 89 animals were selected showing reproductive disorders like abortion, repeat breeding, anoestrous, pyometra, metritis, retention of placenta. No animal had the history of vaccination against brucellosis.

Serological analysis

Three different diagnostic techniques i.e. RBPT,
indirect enzyme linked immunosorbent assay (I-ELISA) and PCR were comparatively evaluated for diagnosis of bovine brucellosis. Rose Bengal Plate Test was conducted as per standard procedure (Alton et al., 1975). Indirect enzyme linked immunosorbent assay was performed as per the manufacturer’s instructions, using the kit procured from Svanova (Biotech-AB), Uppasala, Sweden. For PCR analysis, the genomic DNA of *Brucella* spp. was extracted from sera samples. Briefly, 500 µL of sera samples were placed in a 1.8 ml Eppendorf tube and centrifuged for 15 min at 15,000 × g. The pellet was resuspended in 200 µL of nuclease free water and processed for extraction of DNA using phenol-chloroform method (Sambrook and Russel, 2001). DNA amplification was performed using primers (5’-GAC GAA CGG AAT TTT TCC AAT CCC-3’ and 3’-TGC CGA TCA CTT AAG GGC CTT CAT-5’) (Bricker and Halling, 1994).

**Statistical analysis**

Serological and molecular tests were compared with each other. Sensitivity, specificity, concordance percentage and the agreement between the tests (kappa statistic) were evaluated (Thrusfield, 2008). Arbitrary benchmarks for observed kappa values as described by Thrusfield (2008) were used for evaluating observed kappa values.

**Results**

Out of 89 samples, 44 were positive by I-ELISA, 18 by RBPT and 21 by PCR (Table 1). Of 21 PCR positive sera samples, 47.73% were obtained from seropositive animals, while one serum sample that was negative by the serological tests but its serum was positive by PCR. The agreement of I-ELISA with RBPT and PCR is depicted in Table 2. There was fair agreement between PCR and ELISA (κ=0.48), while a slight agreement was observed between RBPT and ELISA (κ=0.18); and RBPT and PCR (κ=0.11). Analysis of concordance percentage indicated a higher concordance percentage of 74.16% between PCR and I-ELISA. Different tests viz., RBPT and PCR were compared and sensitivity, specificity, predictive value (positive) and predictive value (negative) of different tests when compared with I-ELISA considering it as “golden standard technique” were calculated (Tables 3 and 4). Sensitivity of PCR was the higher (47.73%) than that of RBPT (29.55%). Specificity of PCR was 100%, while the specificity of RBPT was 88.89%.

**Discussion**

Due to difficulty in bacterial isolation, serological tests viz., RBPT, complement fixation test, buffered plate antigen test, I-ELISA are routinely used for its diagnosis. Thirty-one samples that failed to yield a positive reaction in RBPT were positive in the other serological test (I-ELISA), where these samples showed high optical density (OD). Therefore, false negative reaction in RBPT might be due to prozoning effect, when sera of high antibody titers are tested against it (OIE, 2012).

### Table 1: Outcome of individual tests

<table>
<thead>
<tr>
<th>Test</th>
<th>RBPT</th>
<th>I-ELISA</th>
<th>PCR</th>
</tr>
</thead>
<tbody>
<tr>
<td>Positive</td>
<td>18</td>
<td>44</td>
<td>21</td>
</tr>
<tr>
<td>Negative</td>
<td>71</td>
<td>45</td>
<td>68</td>
</tr>
</tbody>
</table>

### Table 2: Agreement between RBPT, ELISA and PCR for diagnosis of bovine brucellosis

<table>
<thead>
<tr>
<th>Combination of tests</th>
<th>Kappa value</th>
<th>95% confidence interval</th>
<th>Concordance percentage (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Lower limit</td>
<td>Upper limit</td>
</tr>
<tr>
<td>RBPT and ELISA</td>
<td>0.1856</td>
<td>0.0485</td>
<td>0.3229</td>
</tr>
<tr>
<td>PCR and ELISA</td>
<td>0.4801*</td>
<td>0.3238</td>
<td>0.6364</td>
</tr>
<tr>
<td>RBPT and PCR</td>
<td>0.1149*</td>
<td>0.0</td>
<td>0.338</td>
</tr>
</tbody>
</table>

* Slight agreement, and * moderate agreement

### Table 3: Sensitivity and specificity of RBPT for diagnosis of bovine brucellosis in comparison to I-ELISA

<table>
<thead>
<tr>
<th>Statistics</th>
<th>Value (%)</th>
<th>95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sensitivity</td>
<td>29.55</td>
<td>16.76 to 45.20%</td>
</tr>
<tr>
<td>Specificity</td>
<td>88.89</td>
<td>75.95 to 96.29%</td>
</tr>
<tr>
<td>Positive predictive value</td>
<td>72.22</td>
<td>46.52 to 90.31%</td>
</tr>
<tr>
<td>Negative predictive value</td>
<td>56.34</td>
<td>44.05 to 68.09%</td>
</tr>
</tbody>
</table>

### Table 4: Sensitivity and specificity of PCR for diagnosis of bovine brucellosis in comparison to I-ELISA

<table>
<thead>
<tr>
<th>Statistics</th>
<th>Value (%)</th>
<th>95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sensitivity</td>
<td>47.73</td>
<td>32.46 to 63.31%</td>
</tr>
<tr>
<td>Specificity</td>
<td>100.00</td>
<td>92.13 to 100.00%</td>
</tr>
<tr>
<td>Positive predictive value</td>
<td>100.00</td>
<td>83.89 to 100.00%</td>
</tr>
<tr>
<td>Negative predictive value</td>
<td>66.18</td>
<td>53.68 to 77.21%</td>
</tr>
</tbody>
</table>

Fig. 1: PCR assay for detection of brucellosis in dairy animals. Lane M: 100 bp DNA ladder, Lane P: Positive control (*Brucella abortus* vaccine), Lane N: Negative control (foetal calf serum), and Lane 1-10: Cell lysate (serum samples)
Out of 89 samples, 44 were positive by I-ELISA, 18 by RBPT and amplicons of 498 bp (Fig. 1) were detected in 21 samples by PCR (Table 1). The wide variation in number of samples detected as positive by RBPT (18), I-ELISA (44) and PCR (21) could be because of many factors. PCR detects DNA, which may be in low quantity in serum samples even though the antibody titre is quite high. Alternatively, the anti-Brucella antibodies titre of serum may be below detectable level but a sufficient DNA quantity for PCR detection as it has been reported in serum may be below detectable level but a sufficient DNA quantity for PCR detection as it has been reported in serum may be below detectable level but a sufficient DNA quantity for PCR detection as it has been reported in serum may be below detectable level but a sufficient DNA quantity for PCR detection as it has been reported in serum may be below detectable level but a sufficient DNA quantity for PCR detection as it has been reported in serum may be below detectable level but a sufficient DNA quantity for PCR detection as it has been reported in serum may be below detectable level but a sufficient DNA quantity for PCR detection as it has been reported in serum may be below detectable level but a sufficient DNA quantity for PCR detection as it has been reported in serum may be below detectable level but a sufficient DNA 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Conflict of interest

The authors declare that they have no conflict of interest.

References


