

Developmental competence of Dromedary camel oocytes fertilized *in vitro* by frozen-thawed ejaculated and epididymal spermatozoa

Scholkamy, T. H.¹; El-Badry, D. A.² and Mahmoud, K. Gh. M.^{3*}

¹Department of Field Investigations, Animal Reproduction Research Institute, Agriculture Research Center, Giza, Egypt;

²Department of Artificial Insemination and Embryo Transfer, Animal Reproduction Research Institute, Agriculture Research Center, Giza, Egypt; ³Department of Animal Reproduction and Artificial Insemination, National Research Center, Dokki, Giza, Egypt

*Correspondence: K. Gh. M. Mahmoud, Department of Animal Reproduction and Artificial Insemination, National Research Center, Dokki, Giza, Egypt. E-mail: karimamahmoud@yahoo.com

(Received 3 Nov 2015; revised version 6 Aug 2016; accepted 24 Aug 2016)

Summary

The present study aimed to compare the *in vitro* fertilizing capacity of frozen-thawed ejaculated and epididymal spermatozoa in order to standardize the semen preparation protocol for camel *in vitro* fertilization (IVF). Semen samples were collected from 7 Dromedary camels by means of artificial vagina (AV). Ten cauda epididymes were obtained from slaughtered adult camels, isolated, incised and rinsed for obtaining the sperm rich fluid. Ejaculated and epididymal spermatozoa were processed for cryopreservation. Fresh and frozen-thawed ejaculated and epididymal spermatozoa were evaluated for motility, livability, membrane and acrosomal integrities. Frozen-thawed ejaculated and epididymal spermatozoa were used to fertilize camel mature oocytes *in vitro*. The results showed that, the progressive motility of freshly collected epididymal spermatozoa was significantly ($P < 0.05$) higher than ejaculated spermatozoa (49.25 ± 1.75 vs. $38.50 \pm 1.50\%$, respectively). The viability index of epididymal spermatozoa was significantly ($P < 0.05$) higher than that of ejaculated spermatozoa (96.63 ± 2.45 vs. 84.00 ± 4.08 , respectively). The post-thaw acrosome and membrane integrities of epididymal spermatozoa were significantly ($P < 0.05$) higher than those of ejaculated spermatozoa. Morula and blastocyst rates of camel oocytes *in vitro* fertilized by frozen-thawed epididymal spermatozoa (59.4 ± 0.8 , 19.12 ± 0.7 and $10.29 \pm 0.7\%$, respectively) were significantly ($P < 0.05$) higher than those fertilized by frozen-thawed ejaculated spermatozoa (48.27 ± 3.1 , 11.63 ± 1.1 and $5.43 \pm 0.8\%$, respectively). In conclusion, the Dromedary camel frozen epididymal spermatozoa have the capacity to endure cryopreservation, fertilize oocytes and produce embryos *in vitro* better than ejaculated sperm.

Key words: Camel, Ejaculated semen, Epididymal spermatozoa, *In vitro* fertilization

Introduction

The camel is an important livestock species uniquely adapted to hot and arid environments. The interest in developing assisted reproductive technologies and cryopreservation for the conservation of camel genetic resources has recently increased. The epididymal sperm from slaughtered or recently died animals will increase the opportunities to create semen and to establish their use for artificial insemination (AI), *in vitro* fertilization (IVF), or intracytoplasmic insemination (Turri *et al.*, 2013; El-Sayed *et al.*, 2015).

There are only scarce reports about the use of stored ejaculated semen for IVF in dromedaries because of the difficulties in semen collection, the gelatinous nature of ejaculated semen and the lack of suitable extenders for its storage. Keeping these problems in view, the use of epididymal spermatozoa could be an alternative. However, availability of viable and functional spermatozoa during the storage period is a prerequisite for AI and IVF, thus necessitating the need for proper storage conditions to maintain the quality and fertilizing ability of the camel spermatozoa for longer periods (El-Badry *et al.*, 2015).

Collection and freezing of epididymal sperm samples has been successfully performed in different species of domestic animals: bulls (Chaveiro *et al.*, 2015), buffalo (Lambrechts *et al.*, 1999), ram (Kaabi *et al.*, 2003), bucks (Turri *et al.*, 2014), equine (Barker and Gandier, 1957), dogs (Yu and Leibo, 2002), cats (Axner *et al.*, 2004), and camels (El-Badry *et al.*, 2015) with the aim of developing techniques suitable for storage of genetic material from these animals.

Freshly collected (Wani *et al.*, 2005), cooled-stored (Badr and Abdel-Malak, 2010) and frozen-thawed (Abdoon *et al.*, 2013; El-Badry *et al.*, 2015) camel epididymal sperm as well as fresh (Khatir *et al.*, 2007) and frozen-thawed (El-Sayed *et al.*, 2012) ejaculated spermatozoa have been successfully used for *in vitro* production of camel embryos. To the best of our knowledge, there was no available report in which the fertilizing capacity of frozen-thawed epididymal as compared to frozen-thawed ejaculated camel spermatozoa has been investigated. Therefore, the objective of this study was to compare the *in vitro* fertilizing capacity of such spermatozoa in order to standardize the semen preparation protocol for IVF, as an attempt to improve the reproductive efficiency of the Dromedary camel.

Materials and Methods

Chemicals

Chemicals and media were purchased from Sigma (St. Louis, MO) unless otherwise stated.

Collection of camel semen

Male Maghrabi camels ($n=7$), 7-2 years of age and 500-650 kg body weight, with a sound history of fertility, raised at Center of Studies and Development of Camel Production, Marsa Matrouh Governorate, Egypt were used in this study. Each camel received 5 kg concentrate feed mixture, 5 kg rice straw and 10 kg green food (*Alfa alfa*) twice daily, while water was offered *ad libitum*.

Semen was collected during the rutting season (December to April) using bovine artificial vagina (AV; 30 cm long and 5 cm internal diameter; IMV, France). A plastic liner was mounted inside the AV and fixed with two plastic ribbons at both ends. After passing the liner through the AV, 8 cm of cylindrical foam (cut longitudinally) was placed between outer jacket of the AV and liner at the end of the AV. A transparent graded glass water-jacketed Pyrex semen collection tube (IMV, France) was attached to the apex of the internal rubber liner. The AV and the water-jacketed semen vessel were filled with water at temperatures of 40 and 35°C, respectively (Ziapour *et al.*, 2015). Males were presented to a sexually receptive female, which was physically restrained in sternal recumbency. Once the male mounted the female, the prepuce was directed toward the AV opening.

Each ejaculate was diluted with equal volume of Shotor (Niasari-Naslaji *et al.*, 2007) in 50 ml glass bottle with screw cap at 37°C in incubator. Using 5 ml single channel pipette (Eppendorf, Germany) the diluted semen was continuously pipetted for 5 min and the bottles were placed in 100 ml bakerys filled with 37°C water and placed for 30 min in 37°C water bath with shaker (Rexmed AI-008, Taiwan) adjusted at 100 RPM. This, in turn, resulted in marked decrease in the viscosity of semen. Just after liquefaction, individual motility and sperm cell concentration were determined by phase contrast microscope and hemocytometer, respectively.

Collection of epididymal spermatozoa

A total of 10 apparently healthy male Dromedary camels, aged between 7 to 10 years, were enrolled in this study during the rutting season. Camel testes were transported from a local abattoir (Kerdasa Abattoir, Giza) to the laboratory in normal saline solution (NSS). Testes were washed with sterile NSS. The cauda epididymides were isolated, incised longitudinally and rinsed 3-4 times with 2 ml of Brackett and Oliphant (BO) medium in 60 mm petri dishes (Bacto Laboratories, Liverpool, Australia) placed on heated stages (37°C).

Cryopreservation and thawing of ejaculated and epididymal sperm

Fluid rich in spermatozoa collected from cauda

epididymides and ejaculated semen were diluted with Shotor (Tris-based egg yolk extender, Niasari-Naslaji *et al.*, 2007) at 37°C in incubator in an appropriate dilution rate to obtain a final concentration of 40×10^6 sperm cell/ml. Diluted samples were then cooled slowly to 5°C in a cooling cabinet for a period of 1.5 h, loaded in 0.25 ml straws (IMV, France) and placed 4 cm above liquid nitrogen in the vapor phase in a foam box for 15 min before being plunged into the liquid phase (El-Badry *et al.*, 2015). Straws were stored in liquid nitrogen until thawing at 37°C in a water bath for 30 s.

Evaluation of the ejaculated and epididymal spermatozoa

Freshly harvested and frozen-thawed spermatozoa were evaluated for total and progressive motilities under phase-contrast microscope. Sperm viability, abnormalities and acrosomal status were evaluated by a dual staining procedure (Didion *et al.*, 1989). For membrane integrity, the procedure described by Jeyendran *et al.* (1984) was used to determine the percentage of hypo-osmotic swelling (HOS) positive sperm cells in each semen sample.

Collection and maturation of oocytes

Camel ovaries were transported from a local abattoir to the laboratory in a thermo container containing PBS at 30°C. Cumulus oocyte complexes (COCs) were aspirated from follicles 2-mm in diameter using 18-gauge needle attached to 10 ml syringe. After being washed 3 times in PBS, COCs with at least 2-compact layers of cumulus cells and a homogeneous cytoplasm were selected (grade A and B; Figs. 1A and B) and washed 3 times in maturation medium. For maturation, COCs were cultured in 100 μ L droplets of maturation medium (10-15 oocytes per droplet) covered with mineral oil for 30 h (Khatir and Anouassi, 2006) at 38.5°C in 5% CO₂ and humidified air. The maturation medium consisted of TCM-199 supplemented with 0.1 mg/ml L-glutamine, 0.8 mg/ml sodium bicarbonate, 0.25 mg/ml pyruvate, 50 μ g/ml gentamicin, 10 μ g/ml bFSH, 10 μ g/ml bLH and 1 μ g/ml estradiol in addition to 20 ng/ml of epidermal growth factor (Wani and Wernery, 2010).

In vitro fertilization

Frozen-thawed spermatozoa collected by AV and from cauda epididymides were prepared for IVF as described by Niwa and Ohgoda (1988). Briefly, the spermatozoa were washed by centrifugation (800 g for 10 min) in BO medium without bovine serum albumin (BSA) and containing 10 mg/ml heparin and 2.5 mM caffeine (Brackett and Oliphant, 1975). The sperm pellets were diluted with BO medium containing 20 mg/ml BSA to adjust the concentration of spermatozoa to 2.5×10^6 sperm/ml. Matured oocytes were washed 3 times in BO medium containing 10 mg/ml BSA and were introduced into 100 μ L droplets of sperm suspension (about 10-15 oocytes/droplet) under paraffin oil, the spermatozoa and oocytes were co-cultured for 5 h under the same culture conditions (5% CO₂, 38.5°C, 95%

humidity). After that the oocytes were washed in TCM-199 to remove attached spermatozoa. Groups of 10-20 oocytes were again replaced with previously prepared co-culture 100 μ L droplets consisting of TCM-199 + 10% FCS. Oocytes of all groups were *in vitro* cultured for 8 days. The morphological appearance of embryos was evaluated under an inverted microscope (Fig. 1D).

Statistical analysis

Two-way analysis of variance and Duncan's multiple range tests were done for the obtained data after angular transformation of percentages to their corresponding arcsin values (Snedecor and Cochran, 1989). Data were analyzed using the 1984-version of Costat (Ecosoft, Inc., USA), and the level of statistical significance was set at $P \leq 0.05$.

Results

Data regarding the pre-freeze and post-thaw characteristics of spermatozoa collected by AV or from cauda epididymides of Dromedary camels are presented in Table 1. Concerning the pre-freeze and post-thaw sperm livability, no significant differences were found

between epididymal and ejaculated camel spermatozoa. The progressive motility of freshly collected epididymal spermatozoa was significantly ($P < 0.05$) higher than that of ejaculated spermatozoa (49.25 ± 1.75 vs. $38.50 \pm 1.50\%$, respectively). The post-thaw total and progressive motility of epididymal spermatozoa at 0, 1, 2 and 3 h post-thaw tended to be higher than those of ejaculated spermatozoa. The viability index of epididymal spermatozoa was significantly ($P < 0.05$) higher than that of ejaculated spermatozoa (96.63 ± 2.45 vs. 84.00 ± 4.08 , respectively). The percentage of fresh and frozen-thawed epididymal spermatozoa with intact acrosomes were significantly ($P < 0.05$) higher than those of ejaculated spermatozoa. Cryopreservation of ejaculated camel spermatozoa resulted in about 30.0% reduction in the percentage of normal acrosomes. The percentage of frozen-thawed epididymal with intact membranes was significantly ($P < 0.05$) higher than that of ejaculated spermatozoa (44.55 ± 0.84 vs. $40.40 \pm 1.03\%$, respectively).

As shown in Table 2, the cleavage rate and the developmental potentials to morula and blastocyst stages of camel oocytes *in vitro* fertilized by frozen-thawed epididymal spermatozoa (59.4 ± 0.8 , 19.12 ± 0.7 , and

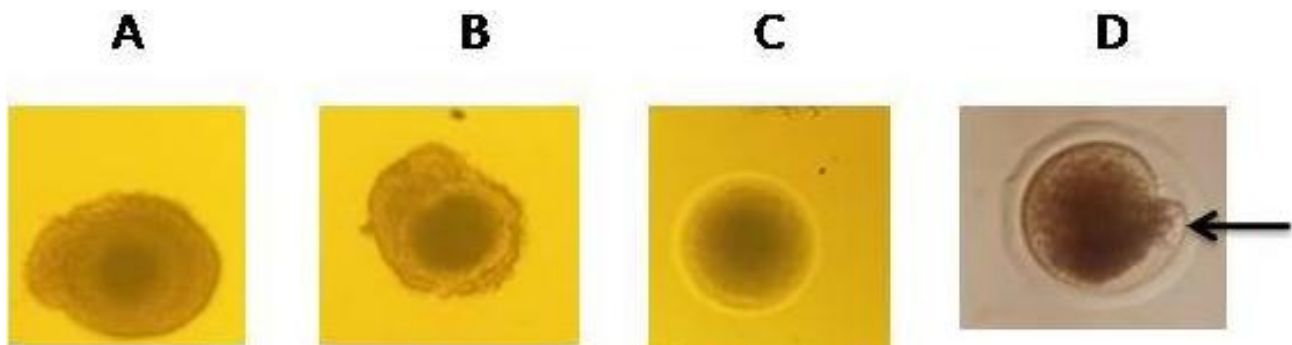


Fig. 1: Immature camel oocytes including grade (A), grade (B) and grade (C) and matured camel oocytes with first polar body extrusion (D)

Table 1: Pre-freeze and post-thaw livability, total and progressive motility of spermatozoa collected by AV or from cauda epididymides of Dromedary camels

Parameters		Semen source	Epididymal	Ejaculated
Livability		Fresh	80.85 ± 1.78^a	78.20 ± 1.07^a
		Frozen	66.85 ± 1.27^a	64.40 ± 2.28^a
Total motility (%)		Fresh	67.40 ± 1.33^a	64.00 ± 1.87^a
Progressive motility (%)			49.25 ± 1.75^a	38.50 ± 1.50^b
Post-thaw total motility (%)	0 h	Frozen	47.25 ± 1.52^a	45.00 ± 3.54^a
	1 h		33.00 ± 1.33^a	29.00 ± 1.87^a
	2 h		26.25 ± 1.24^a	22.50 ± 1.12^a
	3 h		13.75 ± 1.20^a	10.00 ± 2.13^a
Post-thaw progressive motility (%)	0 h		32.50 ± 1.25^a	29.00 ± 1.87^a
	1 h		25.50 ± 1.40^a	21.00 ± 1.87^a
	2 h		18.00 ± 1.11^a	15.00 ± 1.50^a
	3 h		10.50 ± 0.95^a	7.00 ± 1.23^a
Viability index			96.63 ± 2.45^a	84.00 ± 4.08^b
Normal acrosomes (%)		Fresh	87.40 ± 1.43^a	79.00 ± 1.23^b
		Frozen	71.25 ± 0.90^a	56.00 ± 1.92^b
Swollen spermatozoa (HOS +ve %)		Fresh	58.15 ± 1.28^a	57.00 ± 1.14^a
		Frozen	44.55 ± 0.84^a	40.40 ± 1.03^b

Means with different alphabetical superscripts within row are significantly different at $P \leq 0.05$

Table 2: Developmental competence of Dromedary camel oocytes fertilized *in vitro* with frozen-thawed ejaculated and epididymal spermatozoa (mean±SE)

Frozen semen	Total oocytes inseminated	Cleavage No. (%)	Morula No. (%) ^a	Blastocyst No. (%) ^a
Ejaculated	129	60 (48.27 ± 3.1) ^b	15 (11.63 ± 1.1) ^b	7 (5.43 ± 0.8) ^b
Epididymal	136	80 (59.40 ± 0.8) ^a	26 (19.12 ± 0.7) ^a	14 (10.29 ± 0.7) ^a

Means with different alphabetical superscripts within column are significantly different at $P \leq 0.05$

10.29 ± 0.7%, respectively) were significantly ($P < 0.05$) higher than those *in vitro* fertilized by frozen-thawed ejaculated spermatozoa (48.27 ± 3.1, 11.63 ± 1.1, and 5.43 ± 0.8%, respectively).

Discussion

There was no significant differences between the total motility of freshly collected epididymal and ejaculated spermatozoa in the current study (67.4 vs. 64.0%, respectively), but the differences were significant ($P < 0.05$) in case of progressive motility (49.25 vs. 38.50%, respectively). The viscid nature of the ejaculated camel semen and the fact that camel spermatozoa are entrapped in a gel like substance of the seminal plasma might be the reason for low spermatozoa motility (Wani, 2009).

The proportions of totally motile fresh ejaculated spermatozoa (64.00 ± 1.87%) in the present study are higher than 44-61% (Sieme *et al.*, 1990), 30-50% (Billah and Skidmore, 1992), 55% (Hasan *et al.*, 1995) and 56.6 ± 12.7 (Al-Qarawi *et al.*, 2002), while lower than 71-84% (Wani *et al.*, 2008) and 78.3 ± 3.97 (Ziapour *et al.*, 2014) reported in ejaculated semen of Dromedary camel. The proportions of progressively motile spermatozoa (38.50 ± 1.50%) in the present study are lower than 51.2 ± 10.2 (Al-Qarawi *et al.*, 2002) and 44.3 ± 6.41 (Ziapour *et al.*, 2014) reported in ejaculated semen of the same species. These differences could be attributed to individual variation, different extenders, method of collection and liquefaction of semen, season, nutritional status, ... etc.

The post-thaw motility of epididymal spermatozoa was slightly higher (but not significant) and their viability indices were significantly ($P < 0.05$) higher than those of ejaculated spermatozoa. Similar findings of higher post-thaw motility of epididymal spermatozoa were recorded in ram (Garcia-Alvarez *et al.*, 2009), stallion (Monteiro *et al.*, 2011) and boar (Rath and Niemann, 1997). Because ejaculated and epididymal samples were not collected from the same camels, it is possible that the different populations in our study might have differed in sperm quality.

In the present study, the post-thaw acrosome and membrane integrities of epididymal spermatozoa were significantly ($P < 0.05$) higher than those of ejaculated spermatozoa. Cryopreservation processes are known as being damaging to the sperm cells, and to compromise the integrity of acrosomal structures (Wakayama and Yanagimachi, 1998). Reyes-Moreno *et al.* (2002) claimed that bovine epididymal epithelium fluid from cauda epididymides was able to protect sperm against oxidative damage. Similar to our findings, Varisli *et al.*

(2009) concluded that acrosomal integrity of ram epididymal sperm was unaffected by chilling stress but the acrosomal integrity of ejaculated sperm was reduced 40% to 50%. Contrarily, Heise *et al.* (2011) reported that, there was no difference in the percentage of viable acrosome intact sperm noted between epididymal sperm (60%) or ejaculated sperm (64%) of the same stallions following cryopreservation.

As compared with previous studies in Dromedary camel, using frozen-thawed epididymal spermatozoa, the cleavage rate recorded herein (59.4%) was more or less similar to that reported by Wani (2009) (43-60%), and higher than other results (15-32%, Nowshari and Wani, 2005; 37.68%, Badr and Abdel-Malak, 2010; 26.8%, Moawad *et al.*, 2011; 25.37%, Fathi *et al.*, 2014) using freshly collected epididymal spermatozoa. In the present study, the cleavage rate of camel oocytes fertilized by frozen-thawed ejaculated spermatozoa (48.27%) was markedly higher than the cleavage rates of 17-20% reported by Abdoon *et al.* (2007), El-Sayed *et al.* (2012, 2015) using frozen-thawed ejaculated camel spermatozoa, and lower than the rates of 64-68% reported by Khatir and Anouassi (2006) and Khatir *et al.* (2007) using fresh ejaculated camel spermatozoa.

Based on the present data, the cleavage rate of camel oocytes fertilized by epididymal spermatozoa was significantly higher than those fertilized by ejaculated spermatozoa. Similar findings were reported in other animal species like bulls (Katska *et al.*, 1996; Chaveiro *et al.*, 2015), goats (Blash *et al.*, 2000) and boars (Rath and Neiman, 1997). In contrast, El-Sayed *et al.* (2015) in camel found no significant differences between the cleavage rates obtained by epididymal and ejaculated sperm. Also, in cattle, Martins *et al.* (2007) recorded lower cleavage rate when using epididymal spermatozoa as compared with the ejaculated ones.

Using the frozen-thawed epididymal spermatozoa, the morula production rate reported herein (19.12%) was more or less similar to the reported results after IVF of Dromedary oocytes with fresh epididymal spermatozoa (21.32%, Badr and Abdel-Malak, 2010; 20.0%, Fathi *et al.*, 2014). Moreover, the use of frozen-thawed ejaculated spermatozoa resulted in 11.63% morula stage which was lower than the rates recorded by Abdoon *et al.* 2007 (24.3%) and El-Sayed *et al.* (2012) (18.10%).

In the current study, the blastocyst production rate was 10.29% when frozen-thawed epididymal spermatozoa were used. Recent reports of using frozen-thawed camel epididymal spermatozoa recorded a blastocyst production rate ranging from 8.3 to 14.8% (Abdoon *et al.*, 2013; El-Badry *et al.*, 2015). Fresh epididymal spermatozoa have been previously used for IVF of Dromedary oocytes with a blastocyst production rate

ranging from 6 to 24% (Nowshari and Wani, 2005; Wani, 2009; Badr and Abdel-Malak, 2010; El-Sayed *et al.*, 2015). Using the frozen-thawed ejaculated spermatozoa, the blastocyst production rate reported herein (5.43%) was within the range reported after IVF of Dromedary oocytes with frozen-thawed ejaculated spermatozoa (4-8%, El-Sayed *et al.*, 2012, 2015) and was markedly lower than the results after IVF of oocytes with freshly ejaculated camel spermatozoa (23-35%, Khatir and Anouassi, 2006; Khatir *et al.*, 2007). The above mentioned variations in the results of *in vitro* developmental potential of camel spermatozoa could be attributed to the method of semen collection, the source of semen, the semen extender, the methods of semen preparation, age of the animals and also the developmental competence of the oocytes or media used to culture zygotes.

In conclusion, it was concluded that the Dromedary camel epididymal spermatozoa have the capacity to endure cryopreservation, fertilize oocytes and produce embryos *in vitro* better than ejaculated sperm

References

- Abdoon, AS; Kandil, OM; Berisha, B; Kliem, H and Schams, D (2007). Morphology of Dromedary camel oocytes and their ability to spontaneous and chemical parthenogenetic activation. *Reprod. Dom. Anim.*, 42: 88-93.
- Abdoon, AS; Kandil, OM; Pizzi, F; Turri, F; El Atrash, A and Sabra, HA (2013). Effect of semen extender on cryopreservation and fertilization rates of Dromedary camel epididymal spermatozoa. Scientific Conference of Camel Research and Production. Khartoum, Sudan, 17th-18th April 2013.
- Al-Qarawi, HA; Abdel-Rahman, SA; El-Mougy, MS and El-Beley, AA (2002). Use of a new computerized system for evaluation of spermatozoal motility and velocity characteristics in relation to fertility levels in Dromedary bulls. *Anim. Reprod. Sci.*, 74: 1-9.
- Axner, E; Hermansson, U and Linde-Forsberg, C (2004). The effect of Equex STM paste and sperm morphology on post-thaw survival of cat epididymal spermatozoa. *Anim. Reprod. Sci.*, 84: 179-191.
- Badr, MR and Abdel-Malak, MG (2010). *In vitro* fertilization and embryo production in Dromedary camel using epididymal spermatozoa. *Global Vet.*, 4: 271-276.
- Barker, CA and Gandier, SCC (1957). Pregnancy in a mare resulting from frozen epididymal spermatozoa. *Can. J. Comp. Med.*, 21: 47-51.
- Billah, M and Skidmore, JA (1992). The collection, evaluation and deep-freezing of Dromedary camel semen. In: *Proceedings of 1st International Camel Conference*. Dubai, UAE, February 2-6. P: 61 (abst.).
- Blash, S; Melican, D and Gavin, W (2000). Cryopreservation of epididymal sperm obtained at necropsy from goats. *Theriogenology*. 54: 899-905.
- Brackett, BG and Oliphant, G (1975). Capacitation of rabbit spermatozoa *in vitro*. *Biol. Reprod.* 12: 260-274.
- Chaveiro, A; Cerqueira, C; Silva, J; Franco, J and Moreira da Silva, F (2015). Evaluation of frozen thawed cauda epididymal sperms and *in vitro* fertilizing potential of bovine sperm collected from the cauda epididymal. *Iran. J. Vet. Res.*, 16: 188-193.
- Didion, BA; Dobrinsky, JR; Giles, JR and Graves, CN (1989). Staining procedure to detect viability and true acrosome reaction in spermatozoa of various species. *Gamete Res.*, 22: 51-57.
- El-Badry, DA; Scholkamy, TH; Anwer, AM and Mahmoud, KGhM (2015). Assessment of freezability and functional integrity of Dromedary camel spermatozoa harvested from caput, corpus and cauda epididymides. *Alexandria J. Vet. Sci.*, 44: 147-158.
- El-Sayed, A; Ashour, G; Kamel, AM and El-Bahrawy, KA (2015). Assessment of embryo production of Dromedary (*Camelus dromedarius*) using two semen sources and two *in vitro* fertilization techniques. *Egypt. J. Anim. Prod.*, 52: 153-160.
- El-Sayed, A; Sayed, HA; El-Hassanein, EE; Murad, H and Barkawi, AH (2012). Effect of epidermal growth factor on *in vitro* production of camel (*Camelus dromedarius*) embryos by using frozen semen. *Egypt. J. Anim. Prod.*, 49: 39-45.
- Fathi, M; Seida, AA; Sobhy, RR; Darwish, GM; Badr, MR and Moawad, AR (2014). Caffeine supplementation during IVM improves frequencies of nuclear maturation and preimplantation development of Dromedary camel oocytes following IVF. *Theriogenology*. 81: 1286-1292.
- García-Álvarez, O; Maroto-Morales, A; Martínez-Pastor, F; Garde, JJ; Ramón, M; Fernández-Santos, MR; Estesio, MC; Pérez-Guzmán, MD and Soler, AJ (2009). Sperm characteristics and *in vitro* fertilization ability of thawed spermatozoa from Black Manchega ram: electro-ejaculation and postmortem collection. *Theriogenology*. 72: 160-168.
- Hassan, MM; Saed, M and Muqtadir, R (1995). Semen collection by artificial vagina and cryopreservation of camel (*Camelus dromedarius*) spermatozoa. *Pak. Vet. J.*, 15: 105-108.
- Heise, A; Thompson, PN and Gerber, D (2011). Influence of seminal plasma on fresh and post-thaw parameters of stallion epididymal spermatozoa. *Anim. Reprod. Sci.*, 123: 192-201.
- Jeyendran, RS; Vander-Ven, HH; Perez-Pelaez, M; Crabo, BG and Zanevld, LJD (1984). Development of an assay to assess the functional integrity of the human sperm membrane and its relationship to other semen characters. *J. Reprod. Fertil.*, 70: 219-228.
- Kaabi, M; Paz, P; Alvarez, M; Anel, E; Boixo, JC; Rouissi, H; Herraez, P and Anel, L (2003). Effect of epididymis handling conditions on the quality of ram spermatozoa recovered post-mortem. *Theriogenology*. 60: 1249-1259.
- Katska, L; Ryńska, B and Smora, Z (1996). Effect of seminal plasma on the *in vitro* fertilizability of bull spermatozoa. *Anim. Reprod. Sci.*, 44: 23-31.
- Khatir, H and Anouassi, A (2006). The first Dromedary (*Camelus dromedarius*) offspring obtained from *in vitro* matured, *in vitro* fertilized and *in vitro* cultured abattoir-derived oocytes. *Theriogenology*. 65: 1727-1736.
- Khatir, H; Anouassi, A and Tibary, A (2007). Quality and developmental ability of Dromedary (*Camelus dromedarius*) embryos obtained by IVM/IVF, *in vivo* matured/IVF or *in vivo* matured/fertilized oocytes. *Reprod. Dom. Anim.*, 42: 263-270.
- Lambrechts, H; van Niekerk, FE; Coetzer, WA; Cloete, SWP and van der Horst, G (1999). The effect of cryopreservation on the survivability, viability and motility of epididymal African buffalo (*Syncerus caffer*) spermatozoa. *Theriogenology*. 52: 1241-1249.
- Martins, CF; Rumpf, R; Pereira, DC and Dode, MN (2007). Cryopreservation of epididymal bovine spermatozoa from

- dead animals and its uses *in vitro* embryo production. Anim. Reprod. Sci., 101: 326-331.
- Moawad, AR; Darwish, GM; Badr, MR and El-Wishy, AB** (2011). *In vitro* fertilization of Dromedary camel (*Camelus dromedaries*) oocytes with epididymal spermatozoa. Reprod. Fertil. Dev., 24: 192-193.
- Monteiro, GA; Papa, FO; Zahn, FS; Dellaqua Jr, JA; Melo, CM; Maziero, RRD; Avanzi, BR; Alvarenga, MA and Guasti, PN** (2011). Cryopreservation and fertility of ejaculated and epididymal stallion sperm. Anim. Reprod. Sci., 127: 197-201.
- Niasari-Naslaji, A; Mosafieri, S; Bahmani, N; Gerami, A; Gharahdaghi, AA; Abarghani, A and Ghanbari, A** (2007). Semen cryopreservation in Bactrian camel (*Camelus bactrianus*) using Shotor diluent: effects of cooling rates and glycerol concentrations. Theriogenology. 68: 618-625.
- Niwa, K and Ohgoda, O** (1988). Synergistic effect of caffeine and heparin on *in vitro* fertilization of cattle oocytes matured in culture. Theriogenology. 30: 733-741.
- Nowshari, MA and Wani, NA** (2005). Camelid embryo development *in vitro*: effect of protein supplementation in maturation medium and subsequent culture in two different media on fertilization and development. Reprod. Fertil. Dev., 17: 276-282.
- Rath, D and Niemann, H** (1997). *In vitro* fertilization of porcine oocytes with fresh and frozen-thawed ejaculated or frozen-thawed epididymal semen obtained from identical boars. Theriogenology. 47: 785-793.
- Reyes-Moreno, C; Boilard, M; Sullivan, R and Sirard, M** (2002). Characterization and identification of epididymal factors that protect ejaculated bovine sperm during *in vitro* storage. Biol. Reprod., 66: 159-166.
- Sieme, I; Merkt, H; Musa, B; Badreldin, H and Willmen, T** (1990). Liquid and deep-freezing preservation of camel semen using different extenders and methods. In: *Proceedings of the workshop "Is it possible to improve the reproductive performance of the camel?"*, Paris, September 10-12, 1990. PP: 273-284.
- Snedecor, GW and Cochran, WG** (1989). *Statistical methods*. 8th Edn., Ames, IA, USA, Iowa State University Press. P: 97.
- Turri, F; Kandil, OM; Abdoon, AS; Sabra, H; El Atrash, A and Pizzi, F** (2013). Conservation of camel genetic resources: epididymal sperm recovery. The Camel Conference@ SOAS. 29 April 2013. PP: 27-32.
- Turri, F; Madeddu, M; Gliozzi, TM; Gandini, G and Pizzi, F** (2014). Effect of testicle postmortem storage on goat frozen-thawed epididymal sperm quality as a tool to improve gene banking in local breeds. Int. J. Anim. Biosci., 8: 440-447.
- Varisli, O; Uguz, C; Agca, C and Agca, A** (2009). Motility and acrosomal integrity comparisons between electro-ejaculated and epididymal ram sperm after exposure to a range of an isosmotic solutions, cryoprotective agents and low temperatures. Anim. Reprod. Sci., 110: 256-268.
- Wakayama, T and Yanagimachi, R** (1998). Development of normal mice from oocytes injected with freeze-dried spermatozoa. Nat. Biotechnol., 16: 639-646.
- Wani, NA** (2009). *In vitro* embryo production in camel (*Camelus dromedarius*) from *in vitro* matured oocytes fertilized with epididymal spermatozoa stored at 4°C. Anim. Reprod. Sci., 111: 69-79.
- Wani, NA; Billah, M and Skidmore, JA** (2008). Studies on liquefaction and storage of ejaculated Dromedary camel (*Camelus dromedarius*) semen. Anim. Reprod. Sci., 109: 309-318.
- Wani, N; Nowshari, M and Wernery, U** (2005). Storage of camel (*Camelus dromedarius*) epididymal spermatozoa in different extenders and at different temperatures. 38th Annu. Meet. Soc. Stud. Reprod., Quebec, Canada, W693.
- Wani, NA and Wernery, U** (2010). *In vitro* maturation of Dromedary (*Camelus dromedarius*) oocytes: effect of different protein supplementations and epidermal growth factor. Reprod. Dom. Anim., 45: e189-e193.
- Yu, I and Leibo, SP** (2002). Recovery of motile, membrane-intact spermatozoa from canine epididymides stored for 8 days at 4°C. Theriogenology. 57: 1179-1190.
- Ziapour, S; Niasari-Naslaji, A; Mirtavousi, M; Keshavarz, M; Kalantari, A and Adel, H** (2014). Semen collection using phantom in Dromedary. Anim. Reprod. Sci., 151: 15-21.